



## Judd Rice Laboratory

at the USC/Norris Comprehensive Cancer Center

[www.histonecode.com](http://www.histonecode.com)

### Propidium Iodide Staining for FACS

Revised by S Houston  
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#### Procedure

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1. Put approximately 500,000 PBS rinsed cells into 0.5 mL PBS.
2. Add this 0.5 mL drop by drop into an eppendorf tube containing 1 mL 100% EtOH. Mix gently.
  - 2.1. This may be kept at 4 degrees for several months.
  - 2.2. If you want to immediately propidium iodide stain, wait 20 minutes after putting cells in EtOH.
3. Spin cells at 600 g for 5 min and get rid of supernatant.
4. Resuspend cells in 1 mL of Propidium Iodide solution.
  - 4.1. For HeLa cells, greater than 20 minutes is sufficient time to be in PI solution.
  - 4.2. For 293T cells and other cells with a lot of RNA, overnight in PI solution is best, to allow time for RNase to chop up RNA.
5. Place samples in FACS tubes.